

foodproof®

Beer Screening 1 LyoKit Ready Reference Guide

Revision A, December 2023

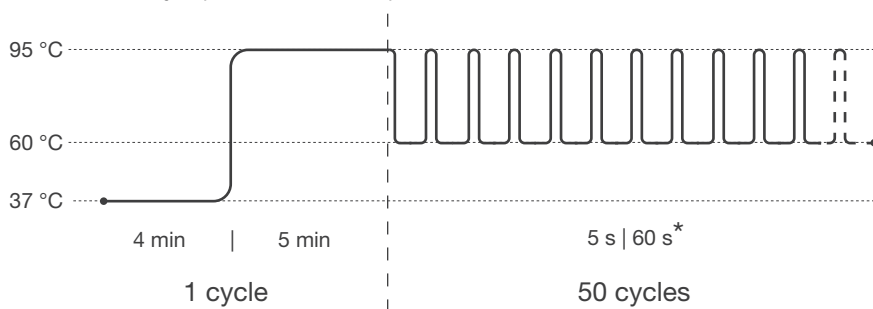
Product No. KIT230071 (LP), KIT230072 (RP)

PCR kit for screening of > 30 beer spoilage bacteria of the genera *Lactobacillus*, *Pediococcus*, *Pectinatus* and *Megasphaera*, including identification of *L. brevis* and detection of hop-tolerance genes *horA* and *horC*. Before starting, it is strongly recommended to read the entire product manual available on our website.

PROGRAM SETUP

Program your real-time PCR instrument before setting up the PCR reactions. Select the following channels:

- ▶ FAM (beer-spoilage bacteria), HEX (*Lactobacillus brevis*), ROX (hop-tolerance related genes) and Cy5 (Internal Control).



Pre-incubation: 1 cycle

Step 1: 37 °C for 4 min

Step 2: 95 °C for 5 min

Amplification: 50 cycles

Step 1 : 95 °C for 5 s

Step 2*: 60 °C for 60 s

* Fluorescence detection

For some real-time PCR instruments the probe quencher as well as the use of a passive reference dye must be specified. This kit contains probes with a non-fluorescent "dark" quencher and no passive reference dye. A Color Compensation is necessary for users of the LightCycler® 480 System: Color Compensation Set 3 (Product No. KIT230005).

DATA INTERPRETATION

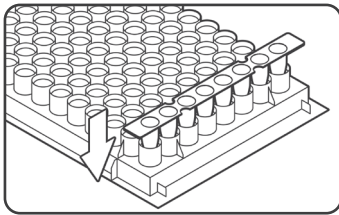
Verify results of positive (Control Template) and negative (H₂O) controls, before interpreting the sample results. Always compare samples to positive and negative controls. Review data from each channel and interpret results as described in the table.

FAM	HEX	ROX	Cy5	Result Interpretation
+	+	+	+ or -	Positive for beer-spoilage bacteria, <i>L. brevis</i> *, hop-tolerance related genes
+	-	+	+ or -	Positive for beer-spoilage bacteria, hop-tolerance related genes
+	+	-	+ or -	Positive for beer-spoilage bacteria, <i>L. brevis</i> *
+	-	-	+ or -	Positive for beer-spoilage bacteria
-	-	+	+ or -	Positive for hop-tolerance related genes
-	-	-	+	Negative for beer-spoilage bacteria, hop-tolerance related genes
-	-	-	-	Invalid

*including *Lactobacillus* sp. DSM 6265 ("*L. brevisimilis*")

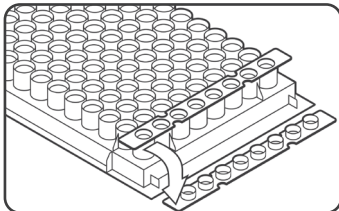
PREPARATION OF THE PCR MIX

Take appropriate precautions to prevent contamination, e.g., by using filter tips and wearing gloves.



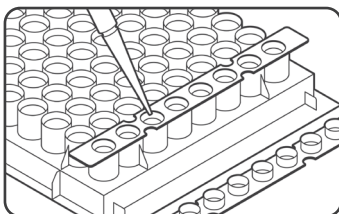
1. PLACE STRIPS IN RACK

Take needed number of PCR tube strips out of aluminum bag. Important: close bag tightly afterwards. Place strips in a suitable PCR tube rack. If needed, gently tap the tubes to move the lyophilized pellets to the bottom of all tubes.



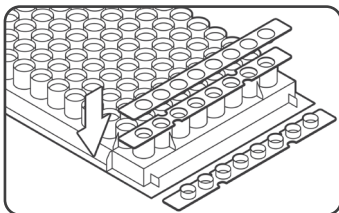
2. DECAP

Immediately before filling, carefully open strips and discard caps. Do not leave open longer than necessary.



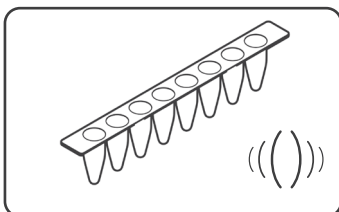
3. ADD SAMPLES AND CONTROLS

Pipette 25 μ L of samples, Negative Control (colorless cap) or Control Template (purple cap) into respective wells. If using less volume, add PCR-grade H₂O to reach 25 μ L.



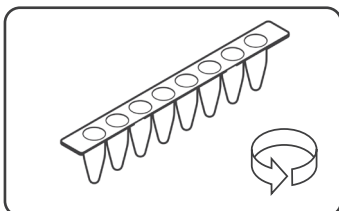
4. SEAL

Carefully seal the tubes with the provided 8-cap strips.



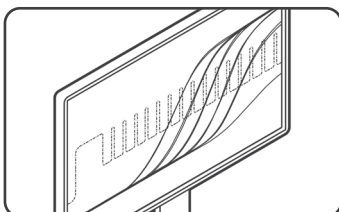
5. MIX

Resuspend pellet after sealing by mixing thoroughly. Alternatively, resuspend pellet by pipetting up and down multiple times in Step 3.



6. CENTRIFUGE

Briefly spin strips, e.g., 5 seconds at 500 - 1,000 x g, in a suitable centrifuge.



7. START REAL-TIME PCR RUN

Cycle samples as described above. Place tubes in a vertical, balanced order into the cycler, e.g., two strips can be placed in the first and last column.